
MiXCR Documentation

Release SNAPSHOT

MiLaboratory

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1	Installation	3
1.1	System requirements	3
1.2	Installation on Mac OS X / Linux using Homebrew	3
1.3	Installation on Mac OS X / Linux / FreeBSD from zip distribution	3
1.4	Installation on Windows	4
2	Quick start	5
2.1	Overview	5
2.2	Basic parameters	5
2.3	Examples	6
2.3.1	Default workflow	6
2.3.2	Analysis of data obtained using 5'RACE-based amplification protocols	7
2.3.3	High quality full length IG repertoire analysis	8
2.3.4	Analysis of RNA-Seq data	9
2.3.5	Assembling of CDR3-based clonotypes for mouse TRB sample	10
2.3.6	Saving links between initial reads and clones	10
3	Alignment	11
3.1	Command line parameters	11
3.2	Aligner parameters	12
3.3	V, J and C aligners parameters	14
3.4	D aligner parameters	15
4	Assemble clones	17
4.1	Command line parameters	18
4.2	Assembler parameters	18
4.3	Separation of clones with same CDR3 (clonal sequence) but different V/J/C genes	19
4.4	Clustering strategy	20
5	Export	21
5.1	Command line parameters	22
5.2	Available fields	22
5.3	Default anchor point positions	24
5.4	Examples	26
5.5	Exporting well formatted alignments for manual inspection	26
5.6	Exporting reads aggregated by clones	27

6	Processing RNA-seq and non-targeted genomic data	29
6.1	Overview	29
6.2	Analysis pipeline	30
6.2.1	Prerequisite	30
6.2.2	Typical analysis workflow	30
6.3	assemblePartial action	31
7	Using external libraries for alignment	33
7.1	IMGT library	33
8	KAligner2: New aligner with big gaps support	35
9	MiXCR Immune Repertoire Analyzer @ BaseSpace®	37
9.1	Input	37
9.2	Parameters	37
9.2.1	Starting material	37
9.2.2	Library type	38
9.2.3	Targeted Library Parameters: 5'-end of the library	38
9.2.4	Targeted Library Parameters: 3'-end of the library	38
9.2.5	Presence of PCR primers and/or adapter sequences	38
9.3	Analysis Settings	38
9.3.1	Target region	38
9.3.2	Filter	38
9.3.3	Output	39
9.4	Hidden parameters	39
9.5	Output	39
10	Gene features and anchor points	41
10.1	Germline features	41
10.1.1	V Gene structure	42
10.1.2	D Gene structure	42
10.1.3	J Gene structure	42
10.2	Mature TCR/BCR gene features	42
10.2.1	V(D)J junction structure	42
10.3	Gene feature syntax	42
10.4	List of predefined gene features	43
10.5	List of predefined reference points	44
11	Appendix	47
11.1	TCR/BCR reference sequences library	47
11.2	Alignment and mutations encoding	47
12	Utility actions	51
12.1	Version info	51
12.2	Merge alignments	51
12.3	Filter alignments	52
13	License	53

MiXCR is a universal framework that processes big immunome data from raw sequences to quantitated clonotypes. MiXCR efficiently handles paired- and single-end reads, considers sequence quality, corrects PCR errors and identifies germline hypermutations. The software supports both partial- and full-length profiling and employs all available RNA or DNA information, including sequences upstream of V and downstream of J gene segments.

MiXCR is free for academic and non-profit use (see [License](#)).

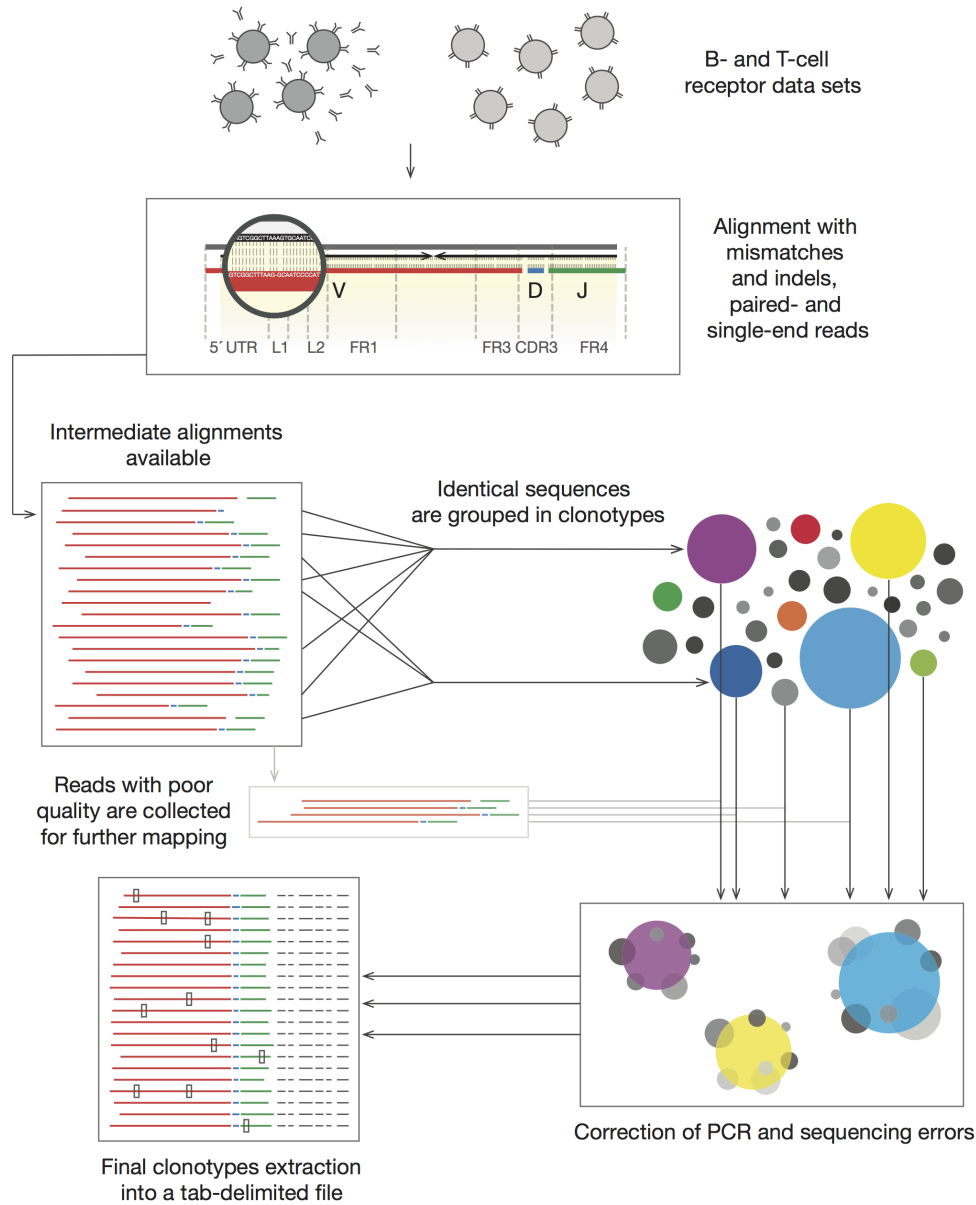


Fig. 1: MiXCR pipeline. The workflow from IG or T-cell receptor data sets to final clonotypes is shown

1.1 System requirements

- Any Java-enabled platform (Windows, Linux, Mac OS X)
- Java version 8 or higher (download from [Oracle web site](#))
- 1–16 Gb RAM (depending on number of clones in the sample)

1.2 Installation on Mac OS X / Linux using Homebrew

[Homebrew](#) is a simple package manager developed for Mac OS X and also [ported](#) to Linux. To install MiXCR using Homebrew just type the following commands:

```
brew tap milaboratory/all
brew install mixcr
```

1.3 Installation on Mac OS X / Linux / FreeBSD from zip distribution

- Check that you have Java **1.8+** installed on your system by typing `java -version`. Here is the example output of this command:

```
> java -version
java version "1.8.0_66"
Java(TM) SE Runtime Environment (build 1.8.0_66-b17)
Java HotSpot(TM) 64-Bit Server VM (build 25.66-b17, mixed mode)
```

- download latest binary distribution of MiXCR from the [release page](#) on GitHub
- unzip the archive

- add extracted folder of MiXCR distribution to your `PATH` variable or add symbolic link for `mixcr` script to your `bin/` folder (e.g. `~/bin/` in Ubuntu and many other popular linux distributions)

1.4 Installation on Windows

Currently there is no execution script or installer for Windows. Still MiXCR can easily be used by direct execution from the jar file.

- check that you have Java **1.7+** installed on your system by typing `java -version`
- download latest binary distribution of MiXCR from the [release page](#) on GitHub
- unzip the archive
- use `mixcr.jar` from the archive in the following way:

```
> java -Xmx4g -Xms3g -jar path_to_mixcr\jar\mixcr.jar ...
```

For example:

```
> java -Xmx4g -Xms3g -jar C:\path_to_mixcr\jar\mixcr.jar align input.fastq.  
↪gz output.vdjca
```

To use `mixcr` from jar file one need to substitute `mixcr` command with `java -Xmx4g -Xms3g -jar path_to_mixcr\jar\mixcr.jar` in all examples from this manual.

2.1 Overview

Typical MiXCR workflow consists of three main processing steps:

- *align*: align sequencing reads to reference V, D, J and C genes of T- or B- cell receptors
- *assemble*: assemble clonotypes using alignments obtained on previous step (in order to extract specific gene regions e.g. CDR3)
- *export*: export alignment (`exportAlignments`) or clones (`exportClones`) to human-readable text file

MiXCR supports the following formats of sequencing data: `fasta`, `fastq`, `fastq.gz`, paired-end `fastq` and `fastq.gz`. As an output of each processing stage, MiXCR produces binary compressed file with comprehensive information about entries produced by this stage (alignments in case of `align` and clones in case of `assemble`). Each binary file can be converted to a human-readable/parsable tab-delimited text file using `exportAlignments` and `exportClones` commands.

2.2 Basic parameters

There are many parameters that user can change to adapt MiXCR for particular needs. While all these parameters are optional there is a set of parameters that are worth considering before running the analysis:

- `-OvParameters.geneFeatureToAlign` sets the gene feature of V gene used for alignment. Applied on the *alignment* stage. Choice of the value for this parameter depends on the type of starting material and library preparation strategy used. There are three options covering most of the cases (see *Gene Features* for the full list):
 - `VRegion` (**default**) is generally suitable for majority of use cases, on the other hand if you have some additional information about your library it is a good idea to use one of the values mentioned below instead of default. Don't change the default value if your library is prepared using multiplex PCR on the V gene side.

- `VTranscript` if RNA was used as a starting material and some kind of non-template-specific technique was used for further amplification on the 5'-end of RNA (e.g. 5'RACE) (see [example](#)). Using of this option is useful for increasing of sequencing information utilization from 5'-end of the molecule, which in turn helps to increase accuracy of V gene identification.
- `VGene` if DNA was used as a starting material and 5' parts of V gene (including V intron, leader sequence and 5'UTR) are supposed to be present in your data. Using of this option is useful for increasing of sequencing information utilization from 5'-end of the molecule, which in turn helps to increase accuracy of V gene identification.

Use `VTranscript` or `VGene` if you plan to assemble full-length clonotypes (including all FRs and CDRs) of T- or B- cell receptors.

- The `-OassemblingFeatures` parameter sets the region of TCR/BCR sequence which will be used to assemble clones. Applied on the [assembly](#) stage. By default its value is `CDR3` which results in assembling of clones by the sequence of *Complementarity Determining Region 3*. To analyse full length sequences use `VDJRegion` as a value for the `assemblingFeatures` (see [Gene Features](#) for more details).
- Another important parameter is `--species`, it sets the target organism. This parameter is used on the [align](#) stage. Possible values are `hsa` (or `HomoSapiens`) and `mmu` (or `MusMusculus`). Default value is `hsa`. This parameter should be supplied on the alignment stage (see [example](#)).

The following sections describes common use cases

2.3 Examples

2.3.1 Default workflow

Tip: Parameters used in this example are particularly suitable for analysis of **multiplex-PCR** selected fragments of T-/B- cell receptor genes.

MiXCR can be used with the default parameters in most cases by executing the following sequence of commands:

```
> mixcr align input_R1.fastq input_R2.fastq alignments.vdjca
... Building alignments
> mixcr assemble alignments.vdjca clones.clns
... Assembling clones
> mixcr exportClones --chains IGH clones.clns clones.txt
... Exporting clones to tab-delimited file
```

The value of only one parameter is changed from its default in this snippet (`--chains IGH`) to tell MiXCR to export only IGH sequences. However even this parameter can be omitted (in this case MiXCR will export all T-/B- cell receptor sequences, that have been found in the sample). We recommend always specify `--chain` parameter at the `exportClones` step to simplify further analysis.

The file produced (`clone.txt`) will contain a tab-delimited table with information about all clonotypes assembled by CDR3 sequence (clone abundance, CDR3 sequence, V, D, J genes, etc.). For full length analysis and other useful features see examples below.

2.3.2 Analysis of data obtained using 5'RACE-based amplification protocols

Consider MiXCR workflow in more detail on analysis of paired-end sequenced cDNA library of IGH gene prepared using 5'RACE-based protocol (i.e. on read covers CDR3 with surroundings and another one covers 5'UTR and downstream sequence of V gene):

1. *Align* raw sequences to reference sequences of segments (V, D, J) of IGH gene:

```
> mixcr align -OvParameters.geneFeatureToAlign=VTranscript \
  --report alignmentReport.log input_R1.fastq input_R2.fastq alignments.vdjca
```

Here we specified non-default value for gene feature used to align V genes (`-OvParameters.geneFeatureToAlign=VTranscript`) in order to utilize information from both reads, more specifically to let MiXCR align V gene's 5'UTRS and parts of coding sequence on 5'-end with sequence from read opposite to CDR3. MiXCR can also produce report file (specified by optional parameter `--report`) containing run statistics which looks like this:

```
Analysis Date: Mon Aug 25 15:22:39 MSK 2014
Input file(s): input_r1.fastq,input_r2.fastq
Output file: alignments.vdjca
Command line arguments: align --report alignmentReport.log input_r1.fastq_
↳input_r2.fastq alignments.vdjca
Total sequencing reads: 323248
Successfully aligned reads: 210360
Successfully aligned, percent: 65.08%
Alignment failed because of absence of V hits: 4.26%
Alignment failed because of absence of J hits: 30.19%
Alignment failed because of low total score: 0.48%
```

One can convert binary output produced by `align` (output.vdjca) to a human-readable text file using `exportAlignments` command.

2. *Assemble* clonotypes:

```
> mixcr assemble --report assembleReport.log alignments.vdjca clones.clns
```

This will build clonotypes and additionally correct PCR and sequencing errors. By default, clonotypes will be assembled by CDR3 sequences; one can specify another gene region by passing additional command line arguments (see *assemble documentation*). The optional report `assembleReport.log` will look like:

```
Analysis Date: Mon Aug 25 15:29:51 MSK 2014
Input file(s): alignments.vdjca
Output file: clones.clns
Command line arguments: assemble --report assembleReport.log alignments.
↳vdjca clones.clns
Final clonotype count: 11195
Total reads used in clonotypes: 171029
Reads used, percent of total: 52.89%
Reads used as core, percent of used: 92.04%
Mapped low quality reads, percent of used: 7.96%
Reads clustered in PCR error correction, percent of used: 0.04%
Clonotypes eliminated by PCR error correction: 72
Percent of reads dropped due to the lack of clonal sequence: 2.34%
Percent of reads dropped due to low quality: 3.96%
Percent of reads dropped due to failed mapping: 5.87%
```

3. *Export* binary file with a list of clones (`clones.clns`) to a human-readable text file:

```
> mixcr exportClones --chains IGH clones.clns clones.txt
```

This will export information about clones with default set of fields, e.g.:

Clone count	Clone fraction	...	V hits	J hits	14. seq. CDR3	AA. seq. CDR3	...
4369	2.9E-3	...	IGHV4-39*00(1388)	IGHJ6*00(131)	TGTGTGAGCCVRHKPM.....		
3477	2.5E-3	...	IGHV4-34*00(1944)	IGHJ4*00(153)	TGTGCGATCAIWDVGL.....		
...

where dots denote text not shown here (for compactness). For the full list of available export options see [export](#) documentation.

Each of the above steps can be customized in order to adapt the analysis pipeline for a specific research task (see below).

2.3.3 High quality full length IG repertoire analysis

For the full length cDNA-based immunoglobulin repertoire analysis we generally recommend to prepare libraries with unique molecular identifiers (UMI) and sequence them using asymmetric paired-end 350 bp + 100 bp Illumina MiSeq sequencing (see [Nature Protocols paper](#)). This approach allows to obtain long-range high quality sequencing and efficiently eliminate PCR and sequencing errors using [MiGEC software](#). The resulting high quality data can be further processed by MiXCR for the efficient full length IGH or IGL repertoire extraction:

1. Merging paired-end reads and *alignment*:

MiXCR's `align` subcommand performs paired-end reads merging and alignment to reference V/D/J and C genes. We recommend using [KAligner2](#) (currently in beta testing) for the full length immunoglobulin profiling:

```
> mixcr align -p kaligner2 -s hsa -r alignmentReport.txt -
↪OreadsLayout=Collinear \
  -OvParameters.geneFeatureToAlign=VTranscript read_R1.fastq.gz read_R2.
↪fastq.gz \
  alignments.vdjca
```

Option `-s` allows to specify species (e.g. homo sapiens - hsa, mus musculus - mmu). Parameter `-OreadsLayout` allow us to set paired-end reads orientation (Collinear, Opposite, Unknown). Note, that after MiGEC analysis paired-end read pairs are in Collinear orientation.

Instead of KAligner2, default MiXCR aligner can be used as well, but it may miss immunoglobulin subvariants that contain several nucleotide-lengths indels within the V gene segment.

2. *Assemble* clones:

```
> mixcr assemble -r assembleReport.txt -OassemblingFeatures=VDJRegion \
  -OseparateByC=true -OqualityAggregationType=Average \
  -OclusteringFilter.specificMutationProbability=1E-5 -
↪OmaxBadPointsPercent=0 \
  alignments.vdjca clones.clns
```

`-OseparateByC=true` separates clones with different antibody isotype.

Set `-OcloneClusteringParameters=null` parameter to switch off the frequency-based correction of PCR errors.

Depending on data quality, one can adjust input threshold by changing the parameter `-ObadQualityThreshold` to improve clonotypes extraction.

See “Assembler parameters” section of documentation for the advanced quality filtering parameters.

3. *Export* clones:

```
> mixcr exportClones -c IGH -o -t clones.clns clones.txt
```

where options `-o` and `-t` filter off the out-of-frame and stop codon containing clonotypes, respectively, and `-c` indicates which chain will be extracted (e.g. IGH, IGL).

2.3.4 Analysis of RNA-Seq data

For detailed description please see [here](#).

MiXCR allows to extract TCR and BCR CDR3 repertoires from RNA-Seq data. Extraction efficiency depends on the abundance of T or B cells in a sample, and also on the sequencing length. 2x150 bp or 2x100 bp paired-end sequencing is recommended. However, even from the paired-end 2x50 bp RNA-Seq data, information on the major clonotypes present (e.g. in a tumor sample) can usually be extracted. The analysis can be performed in the following way:

1. *Align* reads:

```
> mixcr align -p rna-seq -OallowPartialAlignments=true data_R1.fastq.gz data_
↪R2.fastq.gz alignments.vdjca
```

All `mixcr align` parameters can also be used here (e.g. `-s` to specify organism).

`-OallowPartialAlignments=true` option preserves partial alignments for their further use in `assemblePartial`.

2. *Assemble partial* reads:

```
> mixcr assemblePartial alignments.vdjca alignmentsRescued.vdjca
```

To obtain more assembled reads containing full CDR3 sequence it is recommended to perform several iterations of reads assembling using `mixcr assemblePartial` action. `-p` parameter is required for several iterations. In our experience, the best result is obtained after the second iteration:

```
> mixcr assemblePartial alignments.vdjca alignmentsRescued_1.vdjca
> mixcr assemblePartial alignmentsRescued_1.vdjca alignmentsRescued_2.vdjca
```

3. Extend TCR alignments with uniquely determined V and J genes and having incomplete coverage of CDR3s using germline sequences:

```
> mixcr extendAlignments alignmentsRescued_2.vdjca alignmentsRescued_2_
↪extended.vdjca
```

4. *Assemble* clones:

```
> mixcr assemble alignmentsRescued_2_extended.vdjca clones.clns
```

All `mixcr assemble` parameters can also be used here.

- For poor quality data it is recommended to decrease input quality threshold (e.g. `-ObadQualityThreshold=15`).
- To make error correction algorithms to combine clone abundancies add the following option: `-OaddReadsCountOnClustering=true`

5. *Exporting clones:*

```
> mixcr exportClones -c TRA -o -t clones.clns clones.txt
```

One can specify immune receptor chain of interest to extract (`-c TRA` or `-c TRB`, etc) and exclude out-of-frame (option `-o`) and stop codon containing variants (option `-t`).

2.3.5 Assembling of CDR3-based clonotypes for mouse TRB sample

This example shows how to perform routine assembly of clonotypes (based on CDR3 sequence) for mouse TRB library (aligning is performed for all possible genes - TRA/B/D/G and IGH/L/K, but only TRB clones are exported in the final table at the end).

```
> mixcr align --species mmu input_R1.fastq input_R2.fastq alignments.vdjca
```

Other analysis stages can be executed without any additional parameters:

```
> mixcr assemble alignments.vdjca clones.clns
> mixcr exportClones --chains TRB clones.clns clones.txt
```

2.3.6 Saving links between initial reads and clones

In this example we demonstrate how to extract initial read headers for assembled clonotypes. On the `align` step additional `--save-description` option should be specified in order to store headers from reads in the resulting `.vdjca` file:

```
> mixcr align --save-description input_R1.fastq input_R2.fastq alignments.vdjca
```

On the `assemble` stage it is necessary to specify file for the index (which stores mapping from reads to clonotypes):

```
> mixcr assemble --index indexFile alignments.vdjca clones.clns
```

Having this, it is possible to export original read headers with corresponding clone IDs:

```
> mixcr exportAlignments -cloneId indexFile -descrR1 -descrR2 alignments.vdjca_
↪alignments.txt
```

The resulting file `alignments.txt` will look like:

Clone ID	Description R1	Description R2
10	header_1_R1	header_1_R2
	header_2_R1	header_2_R2
2313	header_3_R1	header_3_R2
88142	header_5_R1	header_5_R2
...

The `align` command aligns raw sequencing reads to reference V, D, J and C genes of T- and B- cell receptors. It has the following syntax:

```
mixcr align [options] input_file1 [input_file2] output_file.vdjca
```

MiXCR supports `fasta`, `fastq`, `fastq.gz` and `paired-end fastq` and `fastq.gz` input. In case of `paired-end` reads two input files should be specified.

3.1 Command line parameters

The following table contains description of command line options for `align`:

Option	Default value	Description
-h, --help		Print help message.
-r {file} --report ...		Report file name. If this option is not specified, no report file be produced.
- {chain} --chains ...	ALL	Target immunological chain list separated by “,”. Available values: IGH, IGL, IGK, TRA, TRB, TRG, TRD, IG (for all immunoglobulin chains), TCR (for all T-cell receptor chains), ALL (for all chains) . It is highly recommended to use the default value for this parameter in most cases at the align step. Filtering is also possible at the export step.
-s {speciesName} --species ...	HomoSapiens	Species (organism). Possible values: hsa (or HomoSapiens), mmu (or MusMusculus), rat (currently only TRB, TRA and TRD are supported), or any species from imported IMGT ® library import as described here <i>import segments</i>
-p {parameterName} --parameters ...	default	Preset of parameters. Possible values: default and rna-seq. The rna-seq preset are specifically optimized for analysis of Rna-Seq data (<i>see below</i>)
-t {numberOfThreads} --threads ...	num- ber-of- avail- able CPU cores	Number of processing threads.
-n {numberOfReads} --limit ...		Limit number of sequences that will be analysed (only first -n sequences will be processed from input file(s)).
-a, --save-description		Copy read(s) description line from .fastq or .fasta to .vdjca file (can be then exported with -descrR1 and -descrR2 options in <i>exportAlignments</i> action).
-v, --write-all		Write alignment results for all input reads: including empty results for non-aligned reads.
-g, --save-reads		Copy read(s) from .fastq or .fasta to .vdjca file (this is required for exporting reads aggregated by clones; see <i>this section</i>).
--not-aligned-R1		Write all not aligned reads (R1) to the specified file.
--not-aligned-R2		Write all not aligned reads (R) to the specified file.
-Oparameter=value		Overrides default value of aligner parameter (see next subsection).

All parameters are optional.

3.2 Aligner parameters

MiXCR uses a wide range of parameters that controls aligner behaviour. There are some global parameters and gene-specific parameters organized in groups: vParameters, dParameters, jParameters and cParameters. Each group of parameters may contain further subgroups of parameters etc. In order to override some parameter value one can use -O followed by fully qualified parameter name and parameter value (e.g. -Ogroup1.group2.parameter=value).

One of the key MiXCR features is ability to specify particular *gene regions* which will be extracted from reference

and used as a targets for alignments. Thus, each sequencing read will be aligned to these extracted reference regions. Parameters responsible for target gene regions are:

Parameter	Default value	Description
vParameters.geneFeatureToAlign	VRegion	region in V gene which will be used as target in align
dParameters.geneFeatureToAlign	DRegion	region in D gene which will be used as target in align
jParameters.geneFeatureToAlign	JRegion	region in J gene which will be used as target in align
cParameters.geneFeatureToAlign	CExon1	region in C gene which will be used as target in align

It is important to specify these gene regions such that they will fully cover target clonal gene region which will be used in *assemble* (e.g. CDR3).

One can override default gene regions in the following way:

```
mixcr align -OvParameters.geneFeatureToAlign=VTranscript input_file1 [input_file2]
↳output_file.vdjca
```

Other global aligner parameters are:

Parameter	Default value	Description
allowChimeras	false	Accept alignments with different loci of V and J genes (by default such alignments are dropped).
minSumScore	120	Minimal total alignment score value of V and J genes.
maxHits	5	Maximal number of hits for each gene type: if input sequence align to more than maxHits targets, then only top maxHits hits will be kept.
minimalClonalSequenceLength	12	Minimal clonal sequence length (e.g. minimal sequence of CDR3 to be used for clone assembly)
vjAlignmentOrder (only for single-end analysis)	VThenJ	Order in which V and J genes aligned in target (possible values JThenV and VThenJ). Parameter affects only <i>single-read</i> alignments and alignments of overlapped paired-end reads. Non-overlapping paired-end reads are always processed in VThenJ mode. JThenV can be used for short reads (~100bp) with full (or nearly full) J gene coverage.
relativeMinVFR3CDR3Score (only for paired-end analysis)	7	Relative minimal alignment score of FR3+VCDR3Part region for V gene. V hit will be kept only if its FR3+VCDR3Part part aligns with score greater than relativeMinVFR3CDR3Score * maxFR3CDR3Score, where maxFR3CDR3Score is the maximal alignment score for FR3+VCDR3Part region among all of V hits for current input reads pair.
readsLayout (only for paired-end analysis)	Opposite	Relative orientation of paired reads. Available values: Opposite, Collinear, Unknown.

One can override these parameters in the following way:

```
mixcr align -OmaxHits=3 input_file1 [input_file2] output_file.vdjca
```

3.3 V, J and C aligners parameters

MiXCR uses same types of aligners to align V, J and C genes (KAligner from MiLib; the idea of KAligner is inspired by [this article](#)). These parameters are placed in `parameters` subgroup and can be overridden using e.g. `-OjParameters.parameters.mapperKValue=7`. The following parameters for V, J and C aligners are available:

Parameter	Default V value	Default J value	Default C value	Description
<code>mapperKValue</code>	5	5	5	Length of seeds used in aligner.
<code>floatingLeftBound</code>	false	false	false	Specifies whether left bound of alignment is fixed or float: if <code>floatingLeftBound</code> set to false, the left bound of either target or query will be aligned. Default values are suitable in most cases.
<code>floatingRightBound</code>	false	false	false	Specifies whether right bound of alignment is fixed or float: if <code>floatingRightBound</code> set to false, the right bound of either target or query will be aligned. Default values are suitable in most cases. If your target molecules have no primer sequences in J Region (e.g. library was amplified using primer to the C region) you can change value of this parameter for J gene to <code>false</code> to increase J gene identification accuracy and overall specificity of alignments.
<code>minAlignmentLength</code>	15	15	15	Minimal length of aligned region.
<code>maxAdjacentIndels</code>	2	2	2	Maximum number of indels between two seeds.
<code>absoluteMinScore</code>	40	40	40	Minimal score of alignment: alignments with smaller score will be dropped.
<code>relativeMinScore</code>	0.87	0.87	0.87	Minimal relative score of alignments: if alignment score is smaller than <code>relativeMinScore * maxScore</code> , where <code>maxScore</code> is the best score among all alignments for particular gene type (V, J or C) and input sequence, it will be dropped.
<code>maxHits</code>	7	7	7	Maximal number of hits: if input sequence align with more than <code>maxHits</code> queries, only top <code>maxHits</code> hits will be kept.

These parameters can be overridden like in the following example:

```
mixcr align -OvParameters.parameters.minAlignmentLength=30 \
           -OjParameters.parameters.relativeMinScore=0.7 \
           input_file1 [input_file2] output_file.vdjca
```

Scoring used in aligners is specified by `scoring` subgroup of parameters. It contains the following parameters:

Parameter	Default value	Description
subsMatrix	simple (match = 5, mismatch = -9)	Substitution matrix. Available types: <ul style="list-style-type: none"> • simple — a matrix with diagonal elements equal to match and other elements equal to mismatch • raw — a complete set of 16 matrix elements should be specified; for example: raw(5,-9,-9,-9,-9,5,-9,-9,-9,-9,-9,5,-9,-9,-9,5) (equivalent to the default value)
gapPenalty	-12	Penalty for gap.

Scoring parameters can be overridden in the following way:

```
mixcr align -OvParameters.parameters.scoring.gapPenalty=-20 input_file1 [input_file2] \
↳output_file.vdjca
```

```
mixcr align -OvParameters.parameters.scoring.subsMatrix=simple(match=4,mismatch=-11) \
input_file1 [input_file2] output_file.vdjca
```

3.4 D aligner parameters

The following parameters can be overridden for D aligner:

Parameter	Default value	Description
absoluteMinScore	0	Minimal score of alignment: alignments with smaller scores will be dropped.
relativeMinScore	0.85	Minimal relative score of alignment: if alignment score is smaller than relativeMinScore * maxScore, where maxScore is the best score among all alignments for particular sequence, it will be dropped.
maxHits	3	Maximal number of hits: if input sequence align with more than maxHits queries, only top maxHits hits will be kept.

One can override these parameters like in the following example:

```
mixcr align -OdParameters.absoluteMinScore=10 input_file1 [input_file2] output_file.
↳vdjca
```

Scoring parameters for D aligner are the following:

Parameter	Default value	Description
type	affine	Type of scoring. Possible values: affine, linear.
subsMatrix	simple (match = 5, mismatch = -9)	Substitution matrix. Available types: <ul style="list-style-type: none"> • simple — a matrix with diagonal elements equal to match and other elements equal to mismatch • raw — a complete set of 16 matrix elements should be specified; for example: raw(5, -9, -9, -9, -9, 5, -9, -9, -9, -9, -9, 5, -9, -9, -9, 5) (<i>equivalent to the default value</i>)
gapOpenPenalty	-10	Penalty for gap opening.
gapExtensionPenalty	-1	Penalty for gap extension.

These parameters can be overridden in the following way:

```
mixcr align -OdParameters.scoring.gapExtensionPenalty=-5 input_file1 [input_file2]_
↪output_file.vdjca
```

Assemble clones

The `assemble` command builds a set of clones using alignments obtained with `align` command in order to extract specific gene regions (e.g. CDR3). The syntax of `assemble` is the following:

```
mixcr assemble [options] alignments.vdjca output.clns
```

The following flowchart shows the pipeline of `assemble`:

This pipeline consists of the following steps:

1. The assembler sequentially processes records (aligned reads) from input `.vdjca` file produced by `align`. On the first step, assembler tries to extract gene feature sequences from aligned reads (called *clonal sequence*) specified by `assemblingFeatures` parameter (CDR3 by default); the clonotypes are assembled with respect to *clonal sequence*. If aligned read does not contain clonal sequence (e.g. CDR3 region), it will be dropped.
2. If clonal sequence contains at least one nucleotide with low quality (less than `badQualityThreshold` parameter value), then this record will be deferred for further processing by *mapping procedure*. If fraction of low quality nucleotides in deferred record is greater than `maxBadPointsPercent` parameter value, then this record will be finally dropped. Records with clonal sequence containing only good quality nucleotides are used to build core clonotypes by grouping records by equality of clonal sequences (e.g. CDR3). The sequence quality of the resulting core clonotype will be equal to the total of qualities of the assembled reads. Each core clonotype has two main properties: clonal sequence and `count` — a number of records aggregated by this clonotype.
3. After the core clonotypes are built, MiXCR runs *mapping procedure* that processes records deferred on the previous step. *Mapping* is aimed on rescuing of quantitative information from low quality reads. For this, each deferred record is mapped onto already assembled clonotypes: if there is a fuzzy match, then this record will be aggregated by the corresponding clonotype; in case of several matched clonotypes, a single one will be randomly chosen with weights equal to clonotype counts. If no matches found, the record will be finally dropped.
4. After clonotypes are assembled by initial assembler and mapper, MiXCR proceeds to *clustering*. The clustering algorithm tries to find fuzzy matches between clonotypes and organize matched clonotypes in hierarchical tree (*cluster*), where each child layer is highly similar to its parent but has significantly smaller `count`. Thus, clonotypes with small counts will be attached to highly similar “parent” clonotypes with significantly greater count. The typical cluster looks as follows:

After all clusters are built, only their heads are considered as final clones. The maximal depths of cluster, fuzzy matching criteria, relative counts of parent/childs and other parameters can be customized using `clusteringStrategy` parameters described below.

5. The final step is to align clonal sequences to reference V,D,J and C genes. Since the `assemblingFeatures` are different from those used in `align`, it is necessary to rebuild alignments for clonal sequences. This alignments are built by more accurate aligner (since all hits are known in advance); thus, better alignments will be built for each clonal sequence.
6. The result is written to the binary output file (`.clns`) with a comprehensive information about clones.

4.1 Command line parameters

The command line options of `assemble` are the following:

Option	Default value	Description
<code>-h, --help</code>		Print help message.
<code>-r {file} --report ...</code>		Report file name. If this option is not specified, no report file be produced.
<code>-t {numberOfProcessors} --threads ...</code>	number of available CPU cores	Number of processing threads.
<code>-i {indexFile} --index ...</code>		Specify file which will store information about particular reads aggregated by each clone (mapping readId -> cloneId).
<code>-Oparameter=value</code>		Overrides default value of assembler parameter (see next subsection).

All parameters are optional.

4.2 Assembler parameters

MiXCR uses a wide range of parameters that controls assembler behaviour. There are some global parameters and parameters organized in groups for each stage of assembling: `cloneClusteringParameters` and `cloneFactoryParameters`. Each group of parameters may contain further subgroups of parameters etc. In order to override some parameter value one can use `-O` followed by fully qualified parameter name and parameter value (e.g. `-Ogroup1.group2.parameter=value`).

One of the key MiXCR features is ability to assemble clonotypes by sequence of custom *gene region* (e.g. FR3+CDR3); target clonal sequence can even be disjoint. This region can be specified by `assemblingFeatures` parameter, as in the following example:

```
mixcr assemble -OassemblingFeatures="[V5UTR+L1+L2+FR1,FR3+CDR3]" alignments.vdjca_
↳output.clns
```

(note: `assemblingFeatures` must cover CDR3).

Other global parameters are:

Parameter	Default value	Description
minimalClonality	1	Minimal length of clonal sequence
badQualityThreshold	20	Minimal value of sequencing quality score: nucleotides with lower quality are considered as “bad”. If sequencing read contains at least one “bad” nucleotide within the target gene region, it will be deferred at initial assembling stage, for further processing by mapper.
maxBadPointsPercent	7	Maximal allowed fraction of “bad” points in sequence: if sequence contains more than maxBadPointsPercent “bad” nucleotides, it will be completely dropped and will not be used for further processing by mapper. Sequences with the allowed percent of “bad” points will be mapped to the assembled core clonotypes. Set -OmaxBadPointsPercent=0 in order to completely drop all sequences that contain at least one “bad” nucleotide.
qualityAggregationType	Average	Algorithm used for aggregation of total clonal sequence quality during assembling of sequencing reads. Possible values: Max (maximal quality across all reads for each position), Min (minimal quality across all reads for each position), Average (average quality across all reads for each position), MiniMax (all letters has the same quality which is the maximum of minimal quality of clonal sequence in each read).
minimalQuality		Minimal allowed quality of each nucleotide of assembled clone. If at least one nucleotide in the assembled clone has quality lower than minimalQuality, this clone will be dropped (remember that qualities of reads are aggregated according to selected aggregation strategy during core clonotypes assembly; see qualityAggregationType).
addReadsCountOnClustering	False	Aggregate cluster counts when assembling final clones: if addReadsCountOnClustering is true, then all children clone counts will be added to the head clone; thus head clone count will be a total of its initial count and counts of all its children. Refers to further clustering strategy (see below). Does not refer to mapping of low quality sequencing reads described above.

One can override these parameters in the following way:

```
mixcr assemble -ObadQualityThreshold=10 alignments.vdjca output.clns
```

In order to prevent mapping of low quality reads (filter them off) one can set maxBadPointsPercent to zero:

```
mixcr assemble -OmaxBadPointsPercent=0 alignments.vdjca output.clns
```

4.3 Separation of clones with same CDR3 (clonal sequence) but different V/J/C genes

Since v1.8 MiXCR can separates clones with equal clonal sequence and different V, J and C (e.g. do distinguish clones with different IG isotype) genes.

To make analysis more robust to sequencing errors there is an additional clustering step to shrink artificial diversity generated by this separation mechanism.

The following criteria are used on this pre-clusterization step: more abundant clone (clone1) absorbs smaller clone (clone2) if $clone2.count < clone1.count * maximalPreClusteringRatio$ (cloneX.count denotes number of reads in corresponding clone) and clone2 contain top V/J/C gene from clone1 in it's corresponding gene list.

The following parameter control separation behaviour and pre-clusterization:

Parameter	Default value	Description
maximalPreClusteringRatio	0.0	See conditions for clustering above for more information.
separateByV	false	If false clones with equal clonal sequence but different V gene will be merged into single clone.
separateByJ	false	If false clones with equal clonal sequence but different J gene will be merged into single clone.
separateByC	false	If false clones with equal clonal sequence but different C gene will be merged into single clone.

Example, in order to separate IG clones by isotypes use the following options:

```
mixcr assemble -OseparateByC=true alignments.vdjca output.clns
```

4.4 Clustering strategy

Parameters that control clustering procedure are placed in `cloneClusteringParameters` parameters group which determines the rules for the frequency-based correction of PCR and sequencing errors:

Parameter	Default value	Description
searchDepth	2	Maximum number of cluster layers (not including head).
allowedMutationsInNRegions	Maximum	Maximum allowed number of mutations in N regions (non-template nucleotides in VD, DJ or VJ junctions): if two fuzzy matched clonal sequences will contain more than <code>allowedMutationsInNRegions</code> mismatches in N-regions, they will not be clustered together (one cannot be a direct child of another).
searchParameters	twoMismatchesOrIndels	Parameters that control fuzzy match criteria between clones in adjacent layers. Available predefined values: <code>oneMismatch</code> , <code>oneIndel</code> , <code>oneMismatchOrIndel</code> , <code>twoMismatches</code> , <code>twoIndels</code> , <code>twoMismatchesOrIndels</code> , ... , <code>fourMismatchesOrIndels</code> . By default, <code>twoMismatchesOrIndels</code> allows two mismatches or indels (not more than two errors of both types) between two adjacent clones (parent and direct child).
clusteringFilter	0.0	Probability of a single nucleotide mutation in clonal sequence which has non-hypermutation origin (i.e. PCR or sequencing error). This parameter controls relative counts between two clones in adjacent layers: a smaller clone can be attached to a larger one if its count smaller than count of parent multiplied by $(\text{clonalSequenceLength} * \text{specificMutationProbability}) ^ \text{numberOfMutations}$.

One can override these parameters in the following way:

```
mixcr assemble -OcloneClusteringParameters.searchParameters=oneMismatchOrIndel_
↪alignments.vdjca output.clns
```

In order to turn off clustering one should use the following parameters:

```
mixcr assemble -OcloneClusteringParameters=null alignments.vdjca output.clns
```

Export

In order to export alignment results or clones from a binary file (.vdjca or .clns) to a human-readable text file one can use the `exportAlignments` and `exportClones` commands respectively. The syntax for these commands is:

```
mixcr exportAlignments [options] alignments.vdjca alignments.txt
```

```
mixcr exportClones [options] clones.clns clones.txt
```

The resulting tab-delimited text file will contain columns with different types of information. If no options are specified, the default set of columns - which is sufficient in most cases - will be exported. The possible columns include (see below for details): aligned sequences, qualities, all or just best hit for V, D, J and C genes, corresponding alignments, nucleotide and amino acid sequences of gene region present in sequence, etc. When exporting clones, the additional columns include: clone count, clone fraction etc.

One can customize the list of fields that will be exported by passing parameters to export commands. For example, in order to export just clone count, best hits for V and J genes with corresponding alignments and CDR3 amino acid sequence, one can do:

```
mixcr exportClones -count -vHit -jHit -vAlignment -jAlignment -aaFeature CDR3 clones.  
→clns clones.txt
```

The columns in the resulting file will be exported in exactly the same order as parameters on the command line. The list of available fields will be reviewed in the next subsections. For convenience, MiXCR provides two predefined sets of fields for exporting: `min` (will export minimal required information about clones or alignments) and `full` (used by default); one can use these sets by specifying the `--preset` option:

```
mixcr exportClones --preset min clones.clns clones.txt
```

One can add additional columns to the preset in the following way:

```
mixcr exportClones --preset min -qFeature CDR2 clones.clns clones.txt
```

One can also put all specify export fields in a separate file:

```
-vHits
-dHits
-feature CDR3
...
```

and pass this file to the export command:

```
mixcr exportClones --preset-file myFields.txt clones.clns clones.txt
```

5.1 Command line parameters

The following is a list of command line parameters for both `exportAlignments` and `exportClones`:

Option	Description
<code>-h, --help</code>	print help message
<code>-f, --fields</code>	list available fields that can be exported
<code>-p, --preset</code>	select a predefined set of fields to export (full or min)
<code>-pf,</code> <code>--preset-file</code>	load a file with a list of fields to export
<code>-lf,</code> <code>--list-fields</code>	list available fields that can be exported
<code>-s,</code> <code>--no-spaces</code>	output short versions of column headers to assist with analysis using Pandas, R/DataFrames or another data tables processing library

The following parameters only apply to `exportClones`:

<code>-c, --chains</code>	Limit output to specific locus (e.g. TRA or IGH). Clone fractions will be recalculated accordingly.
<code>-o,</code> <code>--filter-out-of-frames</code>	Exclude out of frames (fractions will be recalculated)
<code>-t, --filter-stops</code>	Exclude sequences containing stop codons (fractions will be recalculated)
<code>-m,</code> <code>--minimal-clone-count</code>	Filter clones by minimal read count.
<code>-q,</code> <code>--minimal-clone-fraction</code>	Filter clones by minimal clone fraction.

5.2 Available fields

The following fields can be exported both for alignments and clones:

Field name	Description
<code>-chains</code>	Chains
<code>-topChains</code>	Chains of top hits
<code>-targets</code>	Number of targets
<code>-vHit</code>	Best V hit
<code>-dHit</code>	Best D hit
<code>-jHit</code>	Best J hit

Field name	Description
-cHit	Best C hit
-vGene	Best V hit gene name (e.g. TRBVA1)
-dGene	Best D hit gene name (e.g. TRBD1)
-jGene	Best J hit gene name (e.g. TRBJ1)
-cGene	Best C hit gene name (e.g. TRBV1)
-vFamily	Best V hit family name (e.g. TRBV)
-dFamily	Best D hit family name (e.g. TRBD)
-jFamily	Best J hit family name (e.g. TRBJ)
-cFamily	Best C hit family name (e.g. TRBV)
-vHitScore	Score for best V hit
-dHitScore	Score for best D hit
-jHitScore	Score for best J hit
-cHitScore	Score for best C hit
-vHitsWithScore	All V hits with score
-dHitsWithScore	All D hits with score
-jHitsWithScore	All J hits with score
-cHitsWithScore	All C hits with score
-vHits	All V hits
-dHits	All D hits
-jHits	All J hits
-cHits	All C hits
-vGenes	All V gene names (e.g. TRBVA1)
-dGenes	All D gene names (e.g. TRBD1)
-jGenes	All J gene names (e.g. TRBJ1)
-cGenes	All C gene names (e.g. TRBV1)
-vFamilies	All V gene family names (e.g. TRBV)
-dFamilies	All D gene family names (e.g. TRBD)
-jFamilies	All J gene family names (e.g. TRBJ)
-cFamilies	All C gene family names (e.g. TRBV)
-vAlignment	Best V alignment
-dAlignment	Best D alignment
-jAlignment	Best J alignment
-cAlignment	Best C alignment
-vAlignments	All V alignments
-dAlignments	All D alignments
-jAlignments	All J alignments
-cAlignments	All C alignments
-nFeature <gene_feature>	Nucleotide sequence of specified gene feature
-qFeature <gene_feature>	Quality string of specified gene feature
-aaFeature <gene_feature>	Amino acid sequence of specified gene feature
-minFeatureQuality <gene_feature>	Minimal quality of specified gene feature
-avrgFeatureQuality <gene_feature>	Average quality of specified gene feature
-lengthOf <gene_feature>	S length of specified gene feature
-nMutations <gene_feature>	Extract nucleotide mutations for specified gene feature
-nMutationsRelative <gene_feature> <relative_to_gene_feature>	Extract nucleotide mutations for specified gene feature relative to another gene feature
-aaMutations <gene_feature>	Extract amino acid mutations for specified gene feature
-aaMutationsRelative <gene_feature> <relative_to_gene_feature>	Extract amino acid mutations for specified gene feature relative to another gene feature
-mutationsDetailed <gene_feature>	Detailed list of nucleotide and amino acid mutations for specified gene feature
-mutationsDetailedRelative <gene_feature> <relative_to_gene_feature>	Detailed list of nucleotide and amino acid mutations for specified gene feature relative to another gene feature

Field name	Description
<code>-positionOf <reference_point></code>	S position of specified reference
<code>-defaultAnchorPoints</code>	Outputs a list of default reference
<code>-vIdentityPercents</code>	V alignment identity percents
<code>-dIdentityPercents</code>	D alignment identity percents
<code>-jIdentityPercents</code>	J alignment identity percents
<code>-cIdentityPercents</code>	C alignment identity percents
<code>-vBestIdentityPercent</code>	Vbest alignment identity percent
<code>-dBestIdentityPercent</code>	Dbest alignment identity percent
<code>-jBestIdentityPercent</code>	Jbest alignment identity percent
<code>-cBestIdentityPercent</code>	Cbest alignment identity percent

The following fields are specific for alignments:

Field name	Description
<code>-readId</code>	Id of read corresponding to alignment
<code>-sequence</code>	Aligned sequence (initial read), or 2 sequences in case of paired-end reads
<code>-quality</code>	Initial read quality, or 2 qualities in case of paired-end reads
<code>-descrR1</code>	Description line from initial .fasta or .fastq file of the first read (only available if <code>-save-description</code> was used in align command)
<code>-descrR2</code>	Description line from initial .fasta or .fastq file of the second read (only available if <code>-save-description</code> was used in align command)
<code>-cloneId <index_file></code>	To which clone alignment was attached.
<code>-cloneIdWithMappingType <index_file></code>	To which clone alignment was attached with additional info on mapping type.

The following fields are specific for clones:

Field name	Description
<code>-cloneId</code>	Unique clone identifier
<code>-count</code>	Clone count
<code>-fraction</code>	Clone fraction
<code>-sequence</code>	Aligned sequence (initial read), or 2 sequences in case of paired-end reads
<code>-quality</code>	Initial read quality, or 2 qualities in case of paired-end reads
<code>-readIds <index_file></code>	Read IDs aggregated by clone.

5.3 Default anchor point positions

Positions of anchor points produced by the `-defaultAnchorPoints` option are outputted as a colon separated list. If an anchor point is not covered by the target sequence nothing is printed for it, but flanking colon symbols are preserved to maintain positions in array. See example:

```
.....:108:117:125:152:186:213:243:244:
```

If there are several target sequences (e.g. paired-end reads or multi-part clonal sequence), an array is outputted for each target sequence. In this case arrays are separated by a comma:

```
2:61:107:107:118:.....,.....:103:112:120:147:181:208:238:239:
```

Even if there are no anchor points in one of the parts:

```
.....,.....:108:117:125:152:186:213:243:244:
```

The following table shows the correspondence between anchor points and positions in the default anchor point array:

Anchors point	Zero-based position	One-based position
V5UTRBeginTrimmed	0	1
V5UTREnd / L1Begin	1	2
L1End / VIntronBegin	2	3
VIntronEnd / L2Begin	3	4
L2End / FR1Begin	4	5
FR1End / CDR1Begin	5	6
CDR1End / FR2Begin	6	7
FR2End / CDR2Begin	7	8
CDR2End / FR3Begin	8	9
FR3End / CDR3Begin	9	10
Number of 3' V deletions (negative value), or length of 3' V P-segment (positive value)	10	11
VEndTrimmed, next position after last aligned nucleotide of V gene	11	12
DBeginTrimmed, position of first aligned nucleotide of D gene	12	13
Number of 5' D deletions (negative value), or length of 5' D P-segment (positive value)	13	14
Number of 3' D deletions (negative value), or length of 3' D P-segment (positive value)	14	15
DEndTrimmed, next position after last aligned nucleotide of D gene	15	16
JBeginTrimmed, position of first aligned nucleotide of J gene	16	17
Number of 3' J deletions (negative value), or length of 3' J P-segment (positive value)	17	18
CDR3End / FR4Begin	18	19
FR4End	19	20
CBegin	20	21
CExon1End	21	22

The following regular expressions can be used to parse the contents of this field in Python:

- for length analysis, or analysis of raw alignments:

```
^(?P<V5UTRBegin>-?[0-9]*):(?P<L1Begin>-?[0-9]*):(?P<VIntronBegin>-?[0-9]*):(?P<L2Begin>-?[0-9]*):(?P<FR1Begin>-?[0-9]*):(?P<CDR1Begin>-?[0-9]*):(?P<FR2Begin>-?[0-9]*):(?P<CDR2Begin>-?[0-9]*):(?P<FR3Begin>-?[0-9]*):(?P<CDR3Begin>-?[0-9]*):(?P<V3Deletion>-?[0-9]*):(?P<VEnd>-?[0-9]*):(?P<DBegin>-?[0-9]*):(?P<D5Deletion>-?[0-9]*):(?P<D3Deletion>-?[0-9]*):(?P<DEnd>-?[0-9]*):(?P<JBegin>-?[0-9]*):(?P<J5Deletion>-?[0-9]*):(?P<CDR3End>-?[0-9]*):(?P<CBegin>-?[0-9]*):(?P<CExon1End>-?[0-9]*)$
```

snipped for Pandas:

```
import pandas as pd
data = pd.read_table("exported.txt", low_memory=False)
anchorPointsRegex="^(?P<V5UTRBegin>-?[0-9]*):(?P<L1Begin>-?[0-9]*):(?P<VIntronBegin>-?[0-9]*):(?P<L2Begin>-?[0-9]*):(?P<FR1Begin>-?[0-9]*):(?P<CDR1Begin>-?[0-9]*):(?P<FR2Begin>-?[0-9]*):(?P<CDR2Begin>-?[0-9]*):(?P<FR3Begin>-?[0-9]*):(?P<CDR3Begin>-?[0-9]*):(?P<V3Deletion>-?[0-9]*):(?P<VEnd>-?[0-9]*):(?P<DBegin>-?[0-9]*):(?P<D5Deletion>-?[0-9]*):(?P<D3Deletion>-?[0-9]*):(?P<DEnd>-?[0-9]*):(?P<JBegin>-?[0-9]*):(?P<J5Deletion>-?[0-9]*):(?P<CDR3End>-?[0-9]*):(?P<CBegin>-?[0-9]*):(?P<CExon1End>-?[0-9]*)$"
#(continues on next page)
```

(continued from previous page)

```
data = pd.concat([data, d.refPoints.str.extract(anchorPointsRegex,
↳expand=True).apply(pd.to_numeric)], axis=1)
```

- A simplified regular expression with a smaller number of fields can be used for analysis of CDR3-assembled clonotypes:

```
^(?:-[0-9]*){8}(?:-[0-9]*):(P<CDR3Begin>-?[0-9]*):(P<V3Deletion>-?[0-9]*):(P<VEnd>-?[0-9]*):(P<DBegin>-?[0-9]*):(P<D5Deletion>-?[0-9]*):(P<D3Deletion>-?[0-9]*):(P<DEnd>-?[0-9]*):(P<JBegin>-?[0-9]*):(P<J5Deletion>-?[0-9]*):(P<CDR3End>-?[0-9]*):(?:-[0-9]*){2}(?:-[0-9]*)
↳$
```

snipped for Pandas:

```
import pandas as pd
data = pd.read_table("exported.txt", low_memory=False)
anchorPointsRegex="^(?:-[0-9]*){8}(?:-[0-9]*):(P<CDR3Begin>-?[0-9]*):(P<V3Deletion>-?[0-9]*):(P<VEnd>-?[0-9]*):(P<DBegin>-?[0-9]*):(P<D5Deletion>-?[0-9]*):(P<D3Deletion>-?[0-9]*):(P<DEnd>-?[0-9]*):(P<JBegin>-?[0-9]*):(P<J5Deletion>-?[0-9]*):(P<CDR3End>-?[0-9]*):(?:-[0-9]*){2}(?:-[0-9]*)$"
data = pd.concat([data, d.refPoints.str.extract(anchorPointsRegex,
↳expand=True).apply(pd.to_numeric)], axis=1)
```

5.4 Examples

Export only the best V, D, J hits and the best V hit alignment from a .vdjca file:

```
mixcr exportAlignments -vHit -dHit -jHit -vAlignment input.vdjca test.txt
```

Best V hit	Best D hit	Best J hit	Best V alignment
IGHV4-34*00		IGHJ4*	00262 452 453 47 237 SC268GSC271ASC275G 956.1,58 303 450 56 301 SG72TSA73CSG136TSA144CSA158CSG171T 331.0
IGHV2-23*00	IGHD2*	IGHJ6*	00262 452 453 47 237 SC268GSC271ASC275G 956.1,58 303 450 56 301 SG72TSA73CSG136TSA144CSA158CSG171T 331.0

The syntax of alignment is described in [appendix](#).

5.5 Exporting well formatted alignments for manual inspection

MiXCR is able to export alignments create with the *align* step as pretty formatted text (human readable) for manual analysis. This can be used both to inspect alignments and to facilitate optimization of analysis parameters and library preparation protocol. To export pretty formatted alignments use the `exportAlignmentsPretty` command:

```
mixcr exportAlignmentsPretty --skip 1000 --limit 10 input.vdjca test.txt
```

this will export 10 results after skipping the first 1000 records, then place the results into the file `test.txt`. Skipping earlier records is often useful because the first sequences in a fastq file may have lower than average read quality.

Omitting the last parameter (output file name) will print results directly to the standard output stream (to console), like this:

```
mixcr exportAlignmentsPretty --skip 1000 --limit 10 input.vdjca
```

Here is a summary of the command line options:

Option	Description
<code>-h, --help</code>	print help message
<code>-n, --limit</code>	limit number of alignments; no more than provided number of results will be outputted
<code>-s, --skip</code>	number of results to skip
<code>-t, --top</code>	output only top hits for V, D, J nad C genes
<code>--cdr3-contains</code>	output only those alignments in which CDR3 contains specified nucleotides (e.g. <code>--cdr3-contains TTCAGAGGAGC</code>)
<code>--read-contains</code>	output only those alignments for which the corresponding reads contain specified nucleotides e.g. <code>--read-contains ATGCTTGCGCGCT</code>)
<code>--verbose</code>	use a more verbose format for alignments (see below for example)

Results produced by this command have the following structure:

Usage of the `--verbose` option will produce alignments in a slightly different format:

5.6 Exporting reads aggregated by clones

MiXCR can preserve the mapping between initial reads and final clonotypes. There are several options for accessing this information.

To enable this behaviour, specify the option `--index` for the *assemble* command:

```
mixcr assemble --index index_file alignments.vdjca output.clns
```

This will tell MiXCR to store an index mapping in the file `index_file` (actually two files will be created: `index_file` and `index_file.p` both of which are used to store the index; in further options one should specify only `index_file` without `.p` extension and MiXCR will automatically read both required files). It is then possible to use this `index_file` in order to map clones to initial reads. For example using the `-cloneId` option for the `exportAlignments` command:

```
mixcr exportAlignments -p min -cloneId index_file alignments.vdjca alignments.txt
```

will print an additional column with the id of the clone which matches the corresponding alignment:

Best V hit	Best D hit	...	CloneId
IGHV4-34*00		...	321
IGHV2-23*00	IGHD2*21	...	
IGHV4-34*00	IGHD2*21	...	22143
...

For more information it's also possible to export the mapping type:

```
mixcr exportAlignments -p min -cloneIdWithMappingType index_file alignments.vdjca ↵
↵alignments.txt
```

Will give you something like:

Best V hit	Best D hit	...	Clone mapping
IGHV4-34*00		...	321:core
IGHV2-23*00	IGHD2*21	...	dropped
IGHV4-34*00	IGHD2*21	...	22143:clustered
IGHV4-34*00	IGHD2*21	...	23:mapped
...

It's also possible to export all read IDs that were aggregated by each clone, using the `-readIds` export options for `exportClones`:

```
mixcr exportClones -c IGH -p min -readIds index_file clones.clns clones.txt
```

This will add a column with a full enumeration of all reads that were clustered into a particular clone:

Clone ID	Clone count	Best V hit	...	Reads
0	7213	IGHV4-34*00	...	56,74,92,96,101,119,169,183...
1	2951	IGHV2-23*00	...	46,145,194,226,382,451,464...
2	2269	IGHV4-34*00	...	58,85,90,103,113,116,122,123...
3	124	IGHV4-34*00	...	240,376,496,617,715,783,813...
...	

Note, the resulting txt file may be *very large* since all read numbers that were successfully assembled will be printed.

Finally, you can export reads aggregated by clone into separate `.fastq` files. To support this behaviour the `align` command needs to be run with the additional `-g` option:

```
mixcr align -g input.fastq alignments.vdjca.gz
```

With this option MiXCR will store the original reads in the `.vdjca` file. Then one can export reads corresponding to a particular clone with the `exportReadsForClones` command. For example, export all reads that were assembled into the first clone (clone with `cloneId = 0`):

```
mixcr exportReadsForClones index_file alignments.vdjca.gz 0 reads.fastq.gz
```

This will create the file `reads_clns0.fastq.gz` (or two files `reads_clns0_R1.fastq.gz` and `reads_clns0_R2.fastq.gz` if the original data were paired) with all reads that were aggregated by the first clone. One can export reads for several clones at a time:

```
mixcr exportReadsForClones index_file alignments.vdjca.gz 0 1 2 33 54 reads.fastq.gz
```

This will create several files (`reads_clns0.fastq.gz`, `reads_clns1.fastq.gz` etc.) for each clone with `cloneId` equal to 0, 1, 2, 33 and 54 respectively.

Processing RNA-seq and non-targeted genomic data

Note: The procedure described here applies to analysis of sequencing data from non-enriched and/or randomly-shred (c)DNA libraries, like [RNA-Seq](#). For analysis of targeted RepSeq data, please see [examples](#) from quick start.

6.1 Overview

Analysis method and quirks described here will also be useful for users who want to extract TCR or Ig repertoire from sequencing data of any other type of non-enriched or randomly shred cDNA / gDNA library.

There are two main challenges of repertoire extraction from non-enriched and randomly-shred libraries:

- **Extraction and alignment of fragments of target molecules.** This procedure must be *sensitive* enough to detect and align sequences with very small parts of V or J genes, but at the same time must be very *selective* not to align non-target sequences homologous to TCR or Ig. Alignment of such sequences and treating them as TCRs or Igs bring a risk of introducing reproducible false-positive clonotypes into resulting clonesets, and may, in turn, lead to detection of false intersections between unlinked repertoires.

MiXCR has a special set of alignment parameters (`-p rna-seq`), which was specifically optimized, and automatically and manually checked on tens of different datasets to give the best possible sensitivity keeping zero false-positive rate.

- **Assembly of overlapping fragmented sequencing reads into long-enough CDR3 containing contigs.** In contrast to sequencing reads from targeted IG or TCR libraries with very determined CDR3 position, reads from randomly shred libraries may cover only a part of CDR3. This fact is especially true for short-read data (like very common 50+50 RNA-Seq), where most part of target sequences only partially cover CDR3. In order to efficiently extract repertoire from such data one have to reconstruct initial CDR3s from fragments scattered all over the initial sequencing dataset. The main challenge of this procedure is, again, the possibility to introduce false-positive clones, namely to perform an overlap between two sequences from different clones. This false positives are not so dangerous as those described in the previous paragraph, but still may introduce certain biases. The problem is that it is very easy to make such false-overlaps as TCR or IG sequences consist mainly from conservative V, D and J regions. So overlapping must be done very carefully, taking into account the positions of all conserved regions.

MiXCR has a special action to perform such an assembly of reads, partially covering CDR3 - `assemblePartial`. Basically it performs an overlap of already aligned reads from `*.vdjca` file, realigns resulting contig, and checks if initial overlap has covered enough part of a non-template N region. Default thresholds in this procedure were optimized to assemble as many contigs as possible while producing zero false overlaps (no false overlaps were detected in all of the benchmarks we have performed).

In case of short reads input, even after `assemblePartial` many contigs/reads still only partially cover CDR3. A substantial fraction of such contigs needs only several nucleotides on the 5' or the 3' end to fill up the sequence up to a complete CDR3. These sequence parts can be taken from the germline, if corresponding V or J gene for the contig is uniquely determined (e.g. from second mate of a read pair). Such procedure is not safe for IGHs, because of hypermutations, but for TCRs which have relatively conservative sequence near conserved Cys and Phe/Trp, it can reconstruct additional clonotypes with relatively small chance to introduce false ones. Described procedure is implemented in the action `extendAlignments`, by default it acts only on TCR sequences.

6.2 Analysis pipeline

MiXCR has all of the steps required to efficiently extract repertoire data from RNA-Seq and similar sequencing datasets, starting from raw `fastq(.gz)` files to final list of clonotypes for each immunological chain (TRB, IGH, etc..).

All default values for analysis parameters were carefully optimized, and should be suitable for most of the use-cases.

6.2.1 Prerequisite

There are only two things you must tell MiXCR for a successful analysis. Both on the first `align` step.

1. **Species.** Using `-s ...` parameter. See [here](#).
2. **Data source origin.** Genomic or transcriptomic. This affects which part of reference V gene sequences will be used for alignment, with or without intron. By default transcriptomic source is assumed, so no additional parameters have to be specified for an analysis of RNA-Seq data. If your data has a genomic DNA origin - e.g. whole genome sequencing (WGS) or whole exome sequencing (WES) - add the following option to the `align` command:

```
-OvParameters.geneFeatureToAlign=VGeneWithP
```

This option tells MiXCR to use unspliced reference sequences of V genes for alignments.

6.2.2 Typical analysis workflow

1. Align sequencing reads against reference V, D, J and C genes.

```
mixcr align -p rna-seq -s hsa -OallowPartialAlignments=true data_R1.fastq.gz_
↪data_R2.fastq.gz alignments.vdjca
```

For single-end data simply specify single input file:

```
mixcr align -p rna-seq -s hsa -OallowPartialAlignments=true data.fastq.gz_
↪alignments.vdjca
```

If your data has a genomic origin add `-OvParameters.geneFeatureToAlign=VGeneWithP` option.

-OallowPartialAlignments=true option is needed to prevent MiXCR from filtering out partial alignments, that don't fully cover CDR3 (the default behaviour while processing targeted RepSeq data). MiXCR will try to assemble contigs using those alignments and reconstruct their full CDR3 sequence on the next step.

2. Perform two rounds of contig assembly (please see [here](#) for available parameters).

```
mixcr assemblePartial alignments.vdjca alignments_rescued_1.vdjca
mixcr assemblePartial alignments_rescued_1.vdjca alignments_rescued_2.vdjca
```

3. (optional) Perform extension of incomplete TCR CDR3s with uniquely determined V and J genes using germline sequences. As described in the *last paragraph of introduction*

```
mixcr extendAlignments alignments_rescued_2.vdjca alignments_rescued_2_
↳extended.vdjca
```

4. Assemble (see [here](#) for details) clonotypes

```
mixcr assemble alignments_rescued_2_extended.vdjca clones.clns
```

5. Export (see [here](#) for details) all clonotypes:

```
mixcr exportClones clones.clns clones.txt
```

or clonotypes for a specific immunological chain:

```
mixcr exportClones -c TRB clones.clns clones.TRB.txt
mixcr exportClones -c IGH clones.clns clones.IGH.txt
...
```

The resulting *.txt files will contain clonotypes along with comprehensive biological information like V, D, J and C genes, clone abundances, etc...

6.3 assemblePartial action

The following options are available for assemblePartial:

Parameter	Default value	Description
kValue	12	Length of k-mer taken from VJ junction region and used for searching potentially overlapping sequences.
kOffset	-7	Offset taken from VEndTrimmed/JBeginTrimmed.
minimalAssembleOverlap	Minimal	Minimal length of the overlapped VJ region: two sequences can be potentially merged only if they have at least minimalAssembleOverlap-wide overlap in the VJ junction region. No mismatches are allowed in the overlapped region.
minimalNOVOverlap	5	Minimal number of non-template nucleotides (N region) that overlap region must cover to accept the overlap.

The above parameters can be specified in e.g. the following way:

```
mixcr assemblePartial -OminimalAssembleOverlap=10 alignments.vdjca alignmentsRescued.
↳vdjca
```

Using external libraries for alignment

Tip: MiXCR utilizes libraries in .json format (see <https://github.com/repseqio> for details).

Note: In some cases when using an external library mixcr will try to establish connection with NCBI over the internet.

7.1 IMGT library

Compiled IMGT library file for MiXCR can be downloaded at <https://github.com/repseqio/library-imgt/releases>. In order to use the library put the .json library file to ~/.mixcr/libraries folder, to the directory from where mixcr is started or to libraries/ subfolder of mixcr installation folder.

Tip: Use `mixcr -v` to see what folders mixcr uses to look for library .json file.

```
> mixcr -v
...
Library search path:
- built-in libraries
- /home/username/.
- /home/username/.mixcr/libraries
- /software/mixcr/libraries
```

```
> mixcr align --library imgt input_R1.fastq input_R2.fastq alignments.vdjca
... Building alignments
```

--library option specifies the library to use for alignment. If the short name is given (ex. "--library imgt") mixcr will look for the latest version in the folder. Otherwise, to use one of the old versions give the full name including the version number (ex. -library imgt.201631-4)

```
> mixcr assemble alignments.vdjca clones.clns
... Assembling clones
> mixcr exportClones --chains IGH clones.clns clones.txt
... Exporting clones to tab-delimited file
```

KAligner2: New aligner with big gaps support

Danger: This feature is provided for beta testing, and not recommended for production use!

To process data using new aligner, apply special parameter pre-sets as follows:

```
mixcr align -p kaligner2 ....
mixcr assemble ....
....
```

Any other parameters can also be provided along with `-p ...` option.

MiXCR Immune Repertoire Analyzer @ BaseSpace®

If your data is deposited at the BaseSpace cloud platform you can perform repertoire extraction and calculate common analysis metrics like V/J usage, without leaving BaseSpace, with the use of MiXCR Immune Repertoire Analyzer app ([this link](#) works if you are logged in into base space, and [this one](#) points to the overview page available without authentication).

9.1 Input

User interface of MiXCR Immune Repertoire Analyzer was specifically optimized to set up best possible analysis pipeline with the minimal possible set of parameters, covering most part of the sequencing data types TCR/IG repertoires can be extracted from.

The list of possible sequencing material sources include, but is not limited to:

- all targeted TCR and IG profiling protocols (both RNA- and gDNA-derived, including multiplex-PCR and 5'RACE based techniques)
- non-enriched RNA-Seq data
- WGS
- single-cell data

9.2 Parameters

9.2.1 Starting material

Sets the type of starting material (RNA or Genomic DNA) of the library. This determines whether MiXCR will look for V intron.

9.2.2 Library type

This option determines whether the data will be treated as amplicon library with the same relative sequence architecture across all reads, where CDR3 is fully covered by each reads. Or, randomly shred library like RNA-Seq or WGS, so MiXCR will perform assembly of target molecules from several reads.

“Random fragments” option works well for shallow libraries like RNA-Seq of solid tissue or sorted cell population, but if the library is too reach with the target molecules (e.g. library was additionally enriched using bait probes), using this option may drastically degrade both computational and “analytical” performance. In this case, select “Targeted TCR/IG library”, no partial CDR3 assembly will be performed, still sequences extracted from reads with full coverage of CDR3, should be enough for the analysis.

9.2.3 Targeted Library Parameters: 5'-end of the library

If you specify the library may contain V primer sequence on the 5' end, and it was not trimmed (see “Presence of PCR primers and/or adapter sequences”), alignment of V genes will not be forcefully extended to the left side, and clonotypes with the same clonal sequence (specified in “Target region”) but different V gene will not be separated into individual records.

Primers for some segments if accidentally annealed to non-target region may introduce chimeric sequences, and prevent exact V gene assignment, thus generating artificial clonotypes differing by only V gene assigned. Splitting by V gene is turned of to prevent this.

9.2.4 Targeted Library Parameters: 3'-end of the library

If you specify the library may contain J or C primer sequence on the 3' end, and it was not trimmed (see “Presence of PCR primers and/or adapter sequences”), alignment of J or C genes respectively will not be forcefully extended to the right side, and clonotypes with the same clonal sequence (specified in “Target region”) but different J or C gene will not be separated into individual records (the motivation is the same as for 5'-end).

9.2.5 Presence of PCR primers and/or adapter sequences

Specifies whether the adapter and primer sequences were trimmed for this data. This applies to V/J/C primers, 5'RACE adapters or may other sequence not originating from target molecules.

It affects V/J/C alignment parameters (see above), and also affects alignment of V 5'UTR sequence (5'UTR will not be aligned if sequence may contain 5'RACE adapter sequences, e.g. if 5'-end of the library do not contain primer sequence is not marked as containing primer, but “Presence of PCR primers and/or adapter sequences” is set to “May be present”).

9.3 Analysis Settings

9.3.1 Target region

Region of the sequence to assemble clones by. Only this part of the sequence will be available in the output.

9.3.2 Filter

Filter out-of-frame CDR3s Output only in-frame reads **Filter out CDR3s with stop codon** Don't output sequences containing stop codons in their target region.

9.3.3 Output

Specify which immunological chains you are interested in. Regardless of the values specified here, data will be aligned against all chains.

9.4 Hidden parameters

All other parameters, like parameters for error correction algorithms, parameters for partial CDR3 reconstruction, extension, etc. are set to their default values, optimal in most cases.

9.5 Output

MiXCR Immune Repertoire Analyzer produces tab separated tables containing comprehensive information about clonotypes detected during analysis. This information includes:

- Clonal sequence
- Aggregated quality score values for clonal sequence
- Anchor positions inside clonal sequence
- Assigned V/D/J/C genes, among with corresponding aggregated alignment scoring
- Encoded alignments of V/J/C genes inside clonal sequence

MiXCR Immune Repertoire Analyzer also contains useful statistics and corresponding charts for the clonesets produced with VDJTools.

Gene features and anchor points

There are several immunologically important parts of TCR/BCR gene (**gene features**). For example, such regions are three complementarity determining regions (CDR1, CDR2 and CDR3), four framework regions (FR1, FR2, FR3 and FR4) etc.

The key feature of MiXCR is the possibility to specify:

- regions of reference V, D, J and C genes sequences that are used in *alignment of raw reads*
- regions of sequence to be exported by *exportAlignments*
- regions of sequence to use as clonal sequence in *clone assembly*
- regions of clonal sequences to be exported by *exportClones*

For convenience, in MiXCR these regions can be specified in terms of above mentioned immunological gene features. The illustrated list of predefined gene features can be found below. The set of possible gene regions is not limited by this list:

- boundary points of gene features (called **anchor points**) can be used to specify begin and end of custom gene regions
- gene features can be concatenated (e.g. VTranscript = {V5UTRBegin:L1End}+{L2Begin:VEnd}).
- offsets can be added or subtracted from original positions of **anchor points** to define even more custom gene regions (for more detailed description see *gene feature syntax*)

Naming of gene features is based on IMGT convention described in *Lefranc et al. (2003), Developmental & Comparative Immunology 27.1 (2003): 55-77*.

10.1 Germline features

Features defined for germline genes are mainly used in *align* and *export*.

10.1.1 V Gene structure

Additionally to core gene features in V region (like FR3) we introduce `VGene`, `VTranscript` and `VRegion` for convenience.

10.1.2 D Gene structure

10.1.3 J Gene structure

10.2 Mature TCR/BCR gene features

Features described here (like `CDR3`) cannot not be used for *align*, since they are not defined for germline genes.

10.2.1 V(D)J junction structure

Important difference between rearranged TCR/BCR sequence and germline sequence of its segments lies in the fact that during V(D)J recombination exact cleavage positions at the end of V gene, begin and end of D gene and begin of J gene varies. As a result in most cases actual `VEnd`, `DBegin`, `DEnd` and `JBegin` anchor positions are not covered by alignment:

In order to use actual V, D, J gene boundaries we introduce four additional anchor positions: `VEndTrimmed`, `DBeginTrimmed`, `DEndTrimmed` and `JBeginTrimmed` and several named gene features: `VDJunction`, `DJJunction` and `VJJunction`. On the following picture one can see the structure of V(D)J junction:

If D gene is not found in the sequence or is not present in target locus (e.g. TRA), `DBeginTrimmed` and `DEndTrimmed` anchor points as well as `VDJunction` and `DJJunction` gene features are not defined.

Similar to `...Trimmed` anchor points in V(D)J junction there is a `V5UTRBeginTrimmed` anchor point representing left bound of alignment upstream start codon. This point is required because 5'UTR could have different length from transcript to transcript, and because library of gene segments inside MiXCR does'n have information on exact 5'UTR lengths.

10.3 Gene feature syntax

Syntax for gene features is the same everywhere. The best way to explain it is by example:

- to enter any gene feature mentioned above or listed in the next section just use its name: `VTranscript`, `CDR2`, `V5UTR` etc.
- to define a gene feature consisting of several concatenated features use `+`: `V5UTR+L1+L2+VRegion` is equivalent to `VTranscript`
- to create gene feature starting at anchor point X and ending at anchor point Y use `{X:Y}` syntax: `{CDR3Begin:CDR3End}` for `CDR3`.

- one can add or subtract offset from original position of anchor point using positive or negative integer value in brackets after anchor point name AnchorPoint(offset): {CDR3Begin(+3):CDR3End} for CDR3 without first three nucleotides (coding conserved cysteine), {CDR3Begin(-6):CDR3End(+6)} for CDR3 with 6 nucleotides downstream its left bound and 6 nucleotides upstream its right bound.
- one can specify offsets for predefined gene feature boundaries using GeneFeatureName(leftOffset, rightOffset) syntax: CDR3(3,0), CDR3(-6,6) - equivalents of two examples from previous item
- all syntax constructs can be combined: {L1Begin(-12):L1End}+L2+VRegion(0,+10).

10.4 List of predefined gene features

Gene Feature Name	Gene feature decomposition	Document
V5UTRGermline	{UTR5Begin:V5UTREnd}	5'UTR; ger
VTranscript	{UTR5Begin:L1End} + {L2Begin:VEnd}	V5UTR + E
VGene	{UTR5Begin:VEnd}	{V5UTRBe
VTranscriptWithP	{UTR5Begin:L1End} + {L2Begin:VEnd} + {VEnd:VEnd(-20)}	V5UTR + E
VGeneWithP	{UTR5Begin:VEnd} + {VEnd:VEnd(-20)}	{V5UTRBe
VDJTranscript	{UTR5Begin:L1End} + {L2Begin:FR4End}	First two ex
V5UTR	{V5UTRBeginTrimmed:V5UTREnd}	5'UTR in a
L1	{L1Begin:L1End}	Part of lider
VLIntronL	{L1Begin:L2End}	L1 + VInt
Exon1	{L1Begin:L1End}	First exon.
L	{L1Begin:L1End} + {L2Begin:L2End}	Full leader
VTranscriptWithout5UTR	{L1Begin:L1End} + {L2Begin:VEnd}	Exon1 + V
VTranscriptWithout5UTRWithP	{L1Begin:L1End} + {L2Begin:VEnd} + {VEnd:VEnd(-20)}	Exon1 + V
VDJTranscriptWithout5UTR	{L1Begin:L1End} + {L2Begin:FR4End}	First two ex
VIntron	{VIntronBegin:VIntronEnd}	Intron in V
L2	{L2Begin:L2End}	Part of lider
Exon2	{L2Begin:FR4End}	Full second
VExon2	{L2Begin:VEnd}	Second exo
VExon2Trimmed	{L2Begin:VEndTrimmed}	Second exo
VRegion	{FR1Begin:VEnd}	Full V Regi
VRegionWithP	{FR1Begin:VEnd} + {VEnd:VEnd(-20)}	Full V Regi
VRegionTrimmed	{FR1Begin:VEndTrimmed}	Full V Regi
FR1	{FR1Begin:FR1End}	Framework
VDJRegion	{FR1Begin:FR4End}	Full V, D, J
CDR1	{CDR1Begin:CDR1End}	CDR1 (Cor
FR2	{FR2Begin:FR2End}	Framework
CDR2	{CDR2Begin:CDR2End}	CDR2 (Cor
FR3	{FR3Begin:FR3End}	Framework
CDR3	{CDR3Begin:CDR3End}	CDR3 (Cor
VCDR3Part	{CDR3Begin:VEndTrimmed}	Part of V re
GermlineVCDR3Part	{CDR3Begin:VEnd}	Part of V re
ShortCDR3	{CDR3Begin(3):CDR3End(-3)}	CDR3 (Cor
VDJunction	{VEndTrimmed:DBeginTrimmed}	N region be
VJunction	{VEndTrimmed:JBeginTrimmed}	Region betv
VPsegment	{VEnd:VEndTrimmed}	P-segment

Gene Feature Name	Gene feature decomposition	Document
GermlineVPSegment	{VEnd:VEnd(-20)}	P-segment
DRegion	{DBegin:DEnd}	Full D Region
DLeftPSegment	{DBeginTrimmed:DBegin}	Left P-segment
DCDR3Part	{DBeginTrimmed:DEndTrimmed}	Full D Region
DJJunction	{DEndTrimmed:JBeginTrimmed}	N region boundary
DRightPSegment	{DEnd:DEndTrimmed}	Right P-segment
GermlineDPSegment	{DEnd:DBegin}	P-segment
DRegionWithP	{DEnd:DBegin} + {DBegin:DEnd} + {DEnd:DBegin}	Full D Region
JRegion	{JBegin:FR4End}	Full J Region
GermlineJCDR3Part	{JBegin:CDR3End}	Part of J region
GermlineJPSegment	{JBegin(20):JBegin}	P-segment
JRegionWithP	{JBegin(20):JBegin} + {JBegin:FR4End}	Full J Region
JPSegment	{JBeginTrimmed:JBegin}	P-segment
JRegionTrimmed	{JBeginTrimmed:FR4End}	Full J Region
JCDR3Part	{JBeginTrimmed:CDR3End}	Part of J region
FR4	{FR4Begin:FR4End}	Framework region
CExon1	{CBegin:CExon1End}	First exon of C region
CRegion	{CBegin:CEnd}	Full C region

10.5 List of predefined reference points

UTR5Begin	Beginning of IG/TCR transcript
V5UTREnd	End of 5'UTR, beginning of IG/TCR CDS as listed in database
V5UTRBeginTrimmed	End of 5'UTR, beginning of IG/TCR CDS as observed in the data
L1Begin	End of 5'UTR, beginning of IG/TCR CDS
L1End	End of first exon, beginning of V intron
VIntronBegin	End of first exon, beginning of V intron
VIntronEnd	End of V intron, beginning of second exon
L2Begin	End of V intron, beginning of second exon
L2End	End of leader sequence, beginning of sequence that codes IG/TCR protein, beginning of FR1.
FR1Begin	End of leader sequence, beginning of sequence that codes IG/TCR protein, beginning of FR1.
FR1End	End of FR1, beginning of CDR1
CDR1Begin	End of FR1, beginning of CDR1
CDR1End	End of CDR1, beginning of FR2
FR2Begin	End of CDR1, beginning of FR2
FR2End	End of FR2, beginning of CDR2
CDR2Begin	End of FR2, beginning of CDR2
CDR2End	End of CDR2, beginning of FR3
FR3Begin	End of CDR2, beginning of FR3
FR3End	End of FR3, beginning of CDR3
CDR3Begin	End of FR3, beginning of CDR3
VEndTrimmed	End of V region after V(D)J rearrangement (commonly inside CDR3)
VEnd	End of V region in genome
DBegin	Beginning of D region in genome
DBeginTrimmed	Beginning of D region after VDJ rearrangement
DEndTrimmed	End of D region after VDJ rearrangement
DEnd	End of D region in genome
JBegin	Beginning of J region in genome

Continued on next page

Table 2 – continued from previous page

JBeginTrimmed	Beginning of J region after V(D)J rearrangement
CDR3End	End of CDR3, beginning of FR4
FR4Begin	End of CDR3, beginning of FR4
FR4End	End of FR4
CBegin	Beginning of C Region
CExon1End	End of C Region first exon (Exon 3 of assembled TCR/IG gene)
CEnd	End of C Region

11.1 TCR/BCR reference sequences library

Default list and sequences of V, D, J and C genes used by MiXCR are taken from GenBank. Accession numbers of records used for each locus are listed in the following table:

<i>Homo sapiens</i>	TRA/TRD	NG_001332.2
	TRB	NG_001333.2
	TRG	NG_001336.2
	IGH	NG_001019.5
	IGK	NG_000834.1
	IGL	NG_000002.1
<i>Mus musculus</i>	TRA/TRD	NG_007044.1
	TRB	NG_006980.1
	TRG	NG_007033.1
	IGH	NG_005838.1
	IGK	NG_005612.1
	IGL	NG_004051.1

11.2 Alignment and mutations encoding

MiXCR outputs alignments in `exportClones` and `exportAlignments` as a list of 7 fields separated by | symbol as follows:

```
targetFrom | targetTo | targetLength | queryFrom | queryTo | mutations |
alignmentScore
```

where

- `targetFrom` - position of first aligned nucleotide in **target sequence** (sequence of gene feature from reference V, D, J or C gene used in alignment; e.g. `VRegion` in TRBV12-2); this boundary is inclusive
- `targetTo` - next position after last aligned nucleotide in **target sequence**; this boundary is exclusive
- `targetLength` - length of **target sequence** (e.g. length of `VRegion` in TRBV12-2)
- `queryFrom` - position of first aligned nucleotide in **query sequence** (sequence of sequencing read or clonal sequence); this boundary is inclusive
- `queryTo` - next position after last aligned nucleotide in **query sequence**; this boundary is exclusive
- `mutations` - list of mutations from **target sequence** to **query sequence** (see below)
- `alignmentScore` - score of alignment

all positions are zero-based (i.e. first nucleotide has index 0)

Mutations are encoded as a list of single-nucleotide edits (similar to what is used in definition of Levenshtein distance, i.e. insertions, deletions or substitutions); if one apply these mutations to aligned subsequence of **target sequence**, one will obtain aligned subsequence of **query sequence**.

Each single mutation (single-nucleotide edit) is encoded in the following way (without any spaces; some fields may absent in some cases, see description):

```
type [fromNucleotide] position [toNucleotide]
```

- type of mutation (one letter):
 - S for substitution
 - D for deletion
 - I for insertion
- `fromNucleotide` is a nucleotide in **target sequence** affected by mutation (applicable only for substitutions and deletions; absent for insertions)
- `position` is a zero-based absolute position in **target sequence** affected by mutation; for insertions denotes position in **target sequence** right after inserted nucleotide
- `toNucleotide` nucleotide after mutation (applicable only for substitutions and insertions; absent for deletions)

Note, that for deletions and substitutions

```
targetSequence[position] == fromNucleotide
```

i.e. target sequence always have `fromNucleotide` at position `position`; for insertions `fromNucleotide` field is absent

Here are several examples of single mutations:

- SA4T - substitution of A at position 4 to T
- DC12 - deletion of C at position 12
- I15G - insertion of G before position 15

Consider the following BLAST-like alignments encoded in MiXCR notation:

- Alignment without mutation

subsequence from `target` (from nucleotide 0 to nucleotide 15) was found to be identical to subsequence from `query` (from nucleotide 3 to nucleotide 18).

- Alignment with mutation

so, to obtain subsequence from **query sequence** from 3 to 18 we need to apply the following mutations to subsequence of **target sequence** from 2 to 16: - deletion of G at position 7 - substitution of C at position 9 to T - insertion of C before at position 13

12.1 Version info

In order to check the current version of MiXCR as usual one can use `-v` option:

```
> mixcr -v
MiXCR v2.1 (built Mon Feb 06 19:56:13 MSK 2017; rev=a9958cd; branch=release/v2.1)
RepSeq.IO v1.2.6 (rev=958e019)
MiLib v1.7.1 (rev=f6ccdbc)
Built-in V/D/J/C library: repseqio.v1.2

Library search path:
- built-in libraries
- /Users/dbolotin/
- /Users/dbolotin/.mixcr/libraries
```

In order to check which version of MiXCR was used to build some `vdjca/clns` file:

```
> mixcr versionInfo file.vdjca
MagicBytes = MiXCR.VDJC.V06
MiXCR v1.8-SNAPSHOT (built Fri Jan 29 16:16:40 MSK 2016; rev=327c30c; branch=feature/
↳mixcr_diff); MiLib v1.2 (rev=4f56782; branch=release/v1.2); MiTools v1.2
↳(rev=eb91603; branch=release/v1.2)
```

12.2 Merge alignments

Allows to merge multiple `.vdjca` files into a single one:

```
> mixcr mergeAlignments file1.vdjca file2.vdjca ... output.vdjca
```

12.3 Filter alignments

Allows to filter alignments in .vdjca file. Example:

```
> mixcr filterAlignments --chains TRA,TRB input_file.vdjca output_file.vdjca
```

The available options are:

Option	Description
-e, --cdr3-equals	Include only those alignments which CDR3 equals to a specified nucleotide sequence
c, --chains	Include only alignments with specified immunological protein chains (comma separated list of some of IGH, IGL, IGK, TRA, TRB, TRG, TRD chains)
-x, --chimeras-only	Output only chimeric alignments
-g, --contains-features	Include only those alignments that contain specified gene feature (see <i>Gene features and anchor points</i>)
-i, --read-ids	Output alignments with specified IDs only
-n, --limit	Maximal number of alignments to process

CHAPTER 13

License

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